

See discussions, stats, and author profiles for this publication at: <https://www.researchgate.net/publication/265690327>

Cloning and expression of gene encoding P23 protein from *Cryptosporidium parvum*

Article · September 2014

CITATIONS

0

READS

177

7 authors, including:



Lan Dinh Thi Bich

Hue University

27 PUBLICATIONS 170 CITATIONS

[SEE PROFILE](#)



Lan Tran Thuy

Hue University

5 PUBLICATIONS 3 CITATIONS

[SEE PROFILE](#)



Viet quoc le

Hue University

4 PUBLICATIONS 4 CITATIONS

[SEE PROFILE](#)



Hoang Tan Quang

Hue University

22 PUBLICATIONS 28 CITATIONS

[SEE PROFILE](#)

Some of the authors of this publication are also working on these related projects:



National Foundation for Science and Technology Development, Vietnam [View project](#)

RESEARCH ARTICLE

Dinh Thi Bich Lan¹
Tran Thuy Lan¹
Le Quoc Viet¹
Phan Van Quyet¹
Hoang Tan Quang¹
Nguyen Hoang Loc²
Phung Thang Long³

Cloning and expression of gene encoding P23 protein from *Cryptosporidium parvum*

Authors' addresses:

¹ Institute of Biotechnology,
Hue University, Hue, Vietnam.

² College of Sciences,
Hue University, Hue, Vietnam.

³ College of Agriculture and
Forestry, Hue University,
Hue, Vietnam.

Correspondence:

Dinh Thi Bich Lan
Institute of Biotechnology,
Hue University, Hue, Vietnam.
Tel.: +84-54-3830208
e-mail: thuhuong@huaf.edu.vn

Article info:

Received: 17 January 2014

Accepted: 6 August 2014

ABSTRACT

We cloned the *cp23* gene coding P23 (glyco)protein from *Cryptosporidium parvum* isolated from Thua Thien Hue province, Vietnam. The coding region of *cp23* gene from *C. parvum* is 99% similar with *cp23* gene deposited in NCBI (accession number: U34390). SDS-PAGE and Western blot analysis showed that the *cp23* gene in *E. coli* BL21 StarTM (DE3) produced polypeptides with molecular weights of approximately 37, 40 and 49 kDa. These molecules may be non-glycosylated or glycosylated P23 fusion polypeptides. Recombinant P23 protein purified by GST (glutathione S-transferase) affinity chromatography can be used as an antigen for *C. parvum* antibody production as well as to develop diagnostic kit for *C. parvum*.

Key words: *cp23* gene, *Cryptosporidium parvum*, P23 protein

Introduction

Cryptosporidium parvum is a protozoan parasite that infects the intestinal epithelium of humans and animals causing the enteric disease. Cryptosporidiosis is a cause of life-threatening disease in immunodeficient people and it has been estimated that 10-15% of patients (or higher) died as a result of infection with *Cryptosporidium* in the world (Jenkins & Fayer, 1995; Wang et al., 2009).

To detect *C. parvum* infection, several methods were developed such as sedimentation-flotation technique (McNabb et al., 1985), oocysts staining (O'Donoghue, 1995), PCR amplification (Laxer et al., 1991). However, these approaches could not be applied widely because of its high cost (Wang et al., 2009). Rapid immuno-chromatographic assays for detecting infections with *C. parvum* in fecal samples were evaluated. The sensitivity (75%) and specificity

(100%) of this assay were relatively high compare with modified Ziehl-Neelsen staining method (Klein et al., 2009; Luginbühl et al., 2005).

A 23 kDa (glyco)protein of *C. parvum* (P23 protein) is a sporozoite surface protein that is geographically conserved among isolated *C. parvum* strains. The P23 protein was identified as an antigen with neutralization-sensitive epitopes. Therefore, it is considered to be a potential candidate for the development of an effective vaccine against cryptosporidiosis, as well as for the diagnosis of cryptosporidiosis by ELISA since it is likely to detect *C. parvum* in various geographical regions (Perryman et al., 1999).

In this work, we report the cloning and expression of gene encoding P23 protein from *C. parvum* isolated in Thua Thien Hue province, Vietnam. Our recombinant P23 protein will be used as an antigen for production of anti-P23 antibody and to

RESEARCH ARTICLE

develop diagnostic kit for *C. parvum* based on immunoassays in the future.

Materials and Methods**Detection of *C. parvum* oocysts**

Bovine fecal samples were collected from barns in Thua Thien Hue province, Vietnam. Sample preparation for detection of *C. parvum* oocysts was performed as described by Wang *et al* (2009).

The oocysts of *C. parvum* were detected by the modified Ziehl-Neelsen (MZN) method. The oocysts stained red on a pale green background and they appear as small discs with 4-6 μm in diameter, but the degree and proportion of staining varies with individual oocysts (World Organisation for Animal Health, 2008).

DNA isolation and amplification of *cp23* gene

Genomic DNA of *C. parvum* was isolated by QIAamp DNA Stool Mini Kit (Qiagen, Max-Volmer-Straße, Hilden). Genomic DNA was then used as template in PCR amplification with the specific primers (Cp23F: 5'-ACGGATCCAAAAATGGGTTGTT-3' and Cp23R: 5'-ACGGATCCCTAATTTAGGCATCA-3'), which designed base on the 23 kDa protein synthetic region of the *cp23* gene from NCBI (accession number: U34390). GGATCC sequence was designed for *Bam*HI cloning site in pGEX 4T-3 expression vector.

The PCR component consisted of 2 μl DNA template, 12.5 μl 2 \times GoTag PCR master mix (Promega, Madison, WI), and 10 pmol each primer. Distilled water was added to a final volume of 25 μl . After 3 min of genomic DNA denaturation at 94°C, 40 cycles of 1 min for denaturation at 94°C, 45 seconds for annealing at 45°C and 1 min for polymerization at 72°C were carried out in the iCycler thermocycler (Bio-Rad, Hercules, CA). In the final cycle, the temperature of 72°C was held for an additional 7 min. The PCR products were purified by Quantum Prep™ Freeze N Squeeze DNA Gel Extraction Spin Columns Kit (Bio-Rad) after electrophoresis on 2% agarose gel.

Cloning *cp23* gene

PCR products (*cp23* gene) were ligated into pGEM®-T Easy vector (Promega), the ligation component consists of 50 ng vector, 5 μl buffer, 3 unit T4 DNA ligase, and 25 ng PCR product. Distilled water was added to a final volume of 10 μl . The ligation was incubated at 25°C for 1 h and at 4°C overnight. Ligation products were transformed into

chemically competent *E. coli* DH5 α cells by heat-shock method. The presence of the insert was determined by colony direct PCR followed by 2% agarose gel electrophoresis. Positive colonies were cultured on 5 ml of LB medium supplemented with 50 $\mu\text{g}/\text{ml}$ ampicillin for biomass production. Recombinant vector (pGEM®-T Easy/*cp23*) was then isolated by PureLink™ Quick Plasmid Miniprep Kit (Invitrogen, Carlsbad, CA). The PCR product was sequenced by the dideoxyterminator method on the 3130 Genetic Analyzer (Applied Biosystems®).

Expression of P23 protein in *E. coli*

The *cp23* gene was cut from pGEM-T Easy/*cp23* vector by *Bam*HI, and purified by Wizard® SV Gel and PCR Clean-Up System Kit (Promega), they were then ligated into *Bam*HI site of pGEX 4T-3 expression vector harboring glutathione S-transferase (GST) gene (GE Healthcare, Björkgatan, Uppsala). The ligation component consists of 0.5 μg vector, 5 μl 2 \times buffer, 4 μg DNA, 3 unit T4 DNA ligase, and distilled water was added to a final volume of 10 μl . The ligation was incubated at 4°C for overnight.

E. coli BL21(DE3) cells containing recombinant pGEX 4T-3/*cp23* vector was cultured in 400 ml LB medium supplemented with 50 $\mu\text{g}/\text{ml}$ ampicillin on a rotation shaker with speed of 200 rpm at 37°C to an OD₆₀₀ value (cell density) of 0.6. Isopropylthio- β -galactoside (IPTG) was added to a final concentration of 1 mM for induction and the culture was then incubated at 25°C for 4 h. The cell biomass was harvested by centrifugation at 10 000 rpm/4°C for 10 min, resuspended in 8 ml TNE buffer (50 mM Tris.HCl pH 7.5, 100 mM NaCl, 2 mM EDTA) and kept on ice bath for 1 hour. Total soluble protein was obtained after sonication for 5 min and centrifugation at 10 000 rpm/4°C for 5 min. Expression levels of P23 protein were determined by sodium dodecyl sulfate-12% polyacrylamide gel electrophoresis (SDS-PAGE, 12% w/v). The gel slice was then stained with Coomassie Blue R-250 and the image was analyzed by Quality One software (ver 4.1, Bio-Rad).

Western blot

Total soluble protein was separated on a SDS-12% PAGE and then electroblotted to a nitrocellulose membrane. Western blot was performed using WesternDot™ 625 Western Blot Kit (Invitrogen) according to the manufacturer's instructions. Anti-P23 antibody was produced in mice by our laboratory (Immunology Laboratory, Institute of Biotechnology, Hue University). The image was collected under UV Transilluminator system (DyNA Light, Labnet,

RESEARCH ARTICLE

Edison, NJ) and analyzed by Quality One software (ver. 4.1, Bio-Rad).

Protein purification

The P23 protein was purified by Glutathione Sepharose™ 4B Purification System (GE Healthcare) according to the manufacturer's instructions. Eight milliliters of cell lysate was added to 2 ml prepared glutathione sepharose 4B and incubated for 30 min at room temperature, the mixture was then transferred to a 15-ml Falcon tube. The glutathione sepharose 4B resin was collected by centrifugation at 500× g for 5 min, washed by adding 5 ml 1× PBS and centrifuged at 500× g for 5 min again, repeat washing step 3 times. P23 protein was eluted by adding 1 ml elution buffer, incubated at room temperature for 30 min and centrifuged at 500× g for 5 min, repeat eluting step 2 times. Eluted protein from each step was mixed to run SDS-PAGE.

Results

Cloning cp23 gene

According to Perryman et al (1996), the cDNA of *cp23* gene has a full-length of 602 bp with a coding region of 336 bp (from nucleotide 111 to 446). The coding frame of *cp23* gene has 345 bp in length including two *Bam*HI ends was amplified by specific primers.

Genomics DNAs from 10 oocysts of *C. parvum* were used as DNA templates for *cp23* gene isolation by PCR amplification. Figure 1 showed bands were of expected size of approximately 350 bp, this size is similar with the coding frame of *cp23* gene.

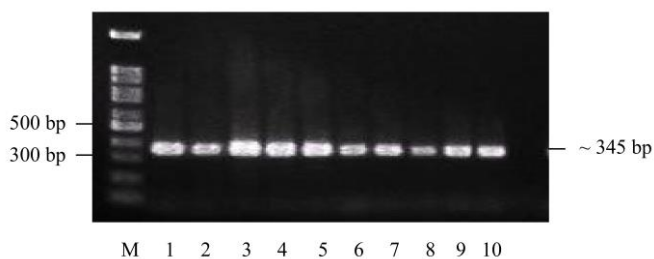


Figure 1. PCR products with specific primers of *cp23* gene. M: DNA size marker (GeneRuler 100 bp DNA Ladder). 1-10: *C. parvum* strains isolated from Thua Thien Hue province.

PCR products were cloned into pGEM-T Easy vector, and one of them was sequenced. The results showed PCR product has 345 bp in length and high similarity with *cp23* gene from

NCBI (99%) (Figure 2). The difference of nucleotide 327 between our *cp23* gene and *cp23* gene from NCBI did not change the deduced amino acid sequence (Figure 3).

```

Query 1 ATGGGTTGTTTCATCATCAAAGCCAGAACTAAAGTTGCTGAAAATAAATCTGCAGCAGAT 60
U34390 1 ATGGGTTGTTTCATCATCAAAGCCAGAACTAAAGTTGCTGAAAATAAATCTGCAGCAGAT 60

Query 61 GCTAACAAACAAGAGAATTAGCTGAAAGAAGGCTCAATTAGCCAAGGCTGTAAGAAT 120
U34390 61 GCTAACAAACAAGAGAATTAGCTGAAAGAAGGCTCAATTAGCCAAGGCTGTAAGAAT 120

Query 121 CCAGCTCCAATCAGCAACCAAGCTCAACAAAAGCCAGAAGAACCAAGAAGTCCGAGCCT 180
U34390 121 CCAGCTCCAATCAGCAACCAAGCTCAACAAAAGCCAGAAGAACCAAGAAGTCCGAGCCT 180

Query 181 GCTCCCAATAATCCTCCAGCTGCTGATGCACCAGCAGCCCAAGCTCCTGCTGCCCTGCT 240
U34390 181 GCTCCCAATAATCCTCCAGCTGCTGATGCACCAGCAGCCCAAGCTCCTGCTGCCCTGCT 240

Query 241 GAACCTGCTCCACAGGATAAGCCAGCTGATGCCCCAGCTGCTGAAGTCCAGCTGCTGAA 300
U34390 241 GAACCTGCTCCACAGGATAAGCCAGCTGATGCCCCAGCTGCTGAAGTCCAGCTGCTGAA 300

Query 301 CCTGCTGCTCAACAAGACAAGCCAGCTGATGCCTAA 336
U34390 301 CCTGCTGCTCAACAAGACAAGCCAGCTGATGCCTAA 336

```

Figure 2. Multiple alignment of the mature peptide region of *cp23* gene from NCBI (accession number: U34390) and that of our *cp23* gene.

```

Query 1 MGCSSSKPETKVAENKSAADANKQRELAEEKKQALAKAVKNPAPISNQAQQKPEEPKSEF
U34390 1 MGCSSSKPETKVAENKSAADANKQRELAEEKKQALAKAVKNPAPISNQAQQKPEEPKSEF

Query 601 APNPNPAADAPAAQAPAPAEPAEPQDKPADAPAAEAPAAEPAQQDKPAD
U34390 60 APNPNPAADAPAAQAPAPAEPAEPQDKPADAPAAEAPAAEPAQQDKPAD

```

Figure 3. Amino acid sequence of mature peptide region of *cp23* gene from NCBI (accession number: U34390) and that of our *cp23* gene.

Expression of cp23 gene

The *cp23* gene was cut from recombinant pGEM T-Easy/*cp3* vector by *Bam*HI and inserted into pGEX 4T-3 expression vector. The pGEX 4T-3/*cp23* vector was transformed into *E. coli* BL21(DE3) cells and positive colonies from selection culture were checked by PCR. Our result showed that a DNA band occurred with the expected size of 345 bp (Figure 4), this suggested that the pGEX 4T-3/*cp23* vector successfully transformed into *E. coli* cells.

Induction was then performed at low temperature of 25°C to avoid formation of inclusion bodies (Makrides, 1996). Expression of the *cp23* gene was expected to produce protein of approximately 49 kDa (23 kDa of P23 protein and 26 kDa of glutathione S-transferase (GST) tag fusion fragment). The results of SDS-PAGE and Western blot showed that there are three protein bands with molecular weights approximate of 49, 40 and 37 kDa (Figure 5).

RESEARCH ARTICLE

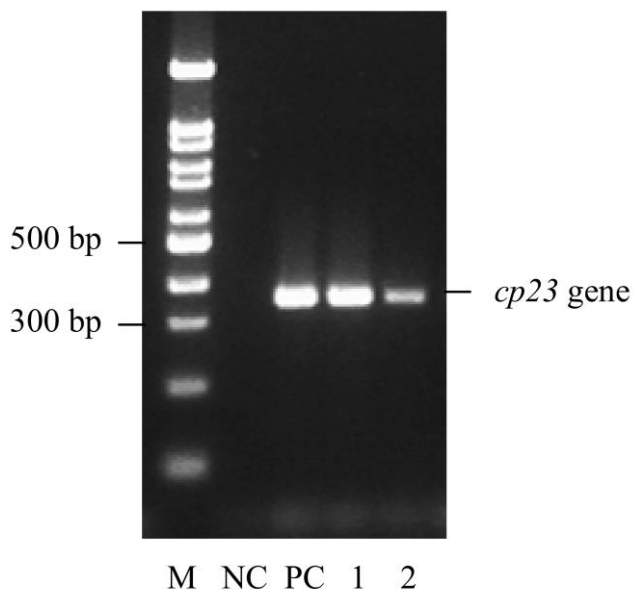


Figure 4. Determination of *cp23* gene in transformed *E. coli* BL21 (DE3) cells. M: DNA size marker, NC: negative control (non-transformed *E. coli* BL21(DE3) cells), PC: positive control (PCR product from pGEM T-Easy/*cp23* vector), 1-2: PCR products from transformed *E. coli* BL21(DE3) cells containing pGEX 4T-3/*cp23* vector.

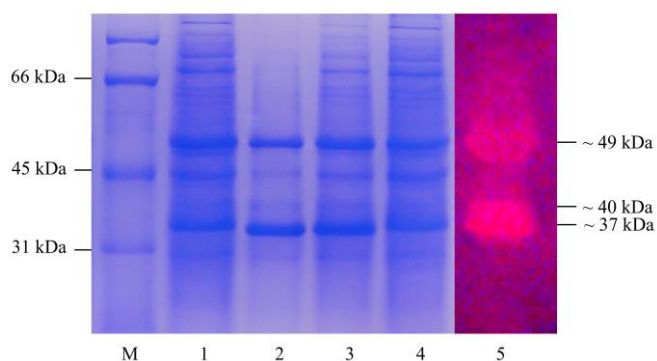


Figure 5. Expression of P23 polypeptides in *E. coli* BL21(DE3) cells. M: protein weight standard, 1-4: SDS-PAGE of P23 fusion polypeptides after IPTG induction from 2-8 h for transformed *E. coli* BL21(DE3) cells, 5: Western blot analysis after 6 h of IPTG induction.

In *C. parvum*, the *cp23* gene encodes a polypeptide chain with 111 amino acids (molecular weight approximate of 11.3 kDa). After P23 protein was synthesized, a *N*-glycosylation was performed to produce a glycoprotein with molecular weight approximate of 23 kDa (or 27 kDa in some cases based on electrophoresis condition) (Priest *et al.*, 1999; Wang

et al., 2009). In this study, the P23 polypeptide chain may be glycosylated or not. If it was glycosylated, molecular weight (fusion protein) will be approximate of 49 kDa or 40 kDa (glycosylated in a different form); if not, molecular weight (fusion polypeptide) will be approximate of 37 kDa (11.3 kDa of P23 polypeptide and 26 kDa of GST tag).

In some previous reports, P23 glycoprotein from *C. parvum* were expressed successfully in *E. coli* based on different expression systems such as pET 30a(+) (Liu *et al.*, 2010), pGEX 4T (Bonafonte *et al.*, 2000), pGEX 4T-2 (Borad *et al.*, 2012; Priest *et al.*, 1999), or pGEX-5X-2 vector (Shahbazi *et al.*, 2009; Shayan *et al.*, 2008). Another host also was used for expression of P23 protein to be *Toxoplasma gondii* (Shirafuji *et al.*, 2005). All of them are glycosylated forms with different molecular weight, i.e. 43 kDa (Shahbazi *et al.*, 2009) or 46 kDa (Bonafonte *et al.*, 2000; Wang *et al.*, 2009) (fusion protein), 23 kDa (Inpankaew *et al.*, 2009; Shahbazi *et al.*, 2009; Shirafuji *et al.*, 2005) or 27 kDa (Liu *et al.*, 2010; Priest *et al.*, 1999) (non-fusion protein). The difference in non- or fusion proteins indicated that P23 protein may be glycosylated in different forms. Other antigenic *C. parvum* protein also expressed in *E. coli* to be CP15 (Jenkins & Fayer, 1995).

The lysate of induced *E. coli* BL21(DE3) cells containing pGEX-4T-3/*cp23* vector were purified by GST chromatography using glutathione sepharose 4B resin. The results of SDS-PAGE showed two strong protein bands have molecular weights of approximately 37 kDa and 49 kDa, another one is very weak (appx. 40 kDa) (Figure 6).

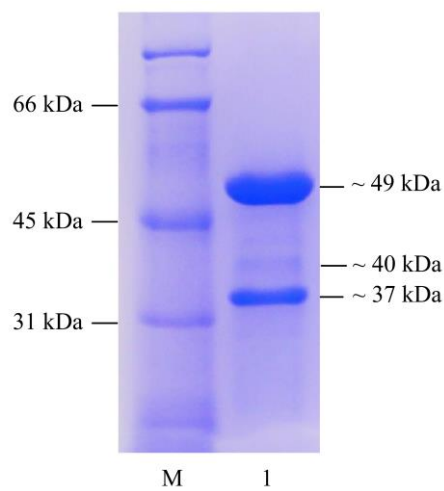


Figure 6. Protein purification by GST chromatography. M: protein weight standard, 1: P23 polypeptides.

RESEARCH ARTICLE

Conclusion

A *cp23* gene coding P23 protein from *C. parvum* was cloned and its sequence has 99% similarity with the *cp23* gene from NCBI. The *cp23* gene in *E. coli* BL21(DE3) produced polypeptides with molecular weights of approximately 37, 40 and 49 kDa; they may be non-glycosylated and glycosylated fusion polypeptides. Western blot analysis showed that 3 fusion polypeptide chains have a GST tag and an epitope for immunological response. Protein purification using GST affinity chromatography was harvested two strong bands (37 and 49 kDa).

Acknowledgement

This study was funded by the Ministry of Science and Technology of Vietnam (No. 02/ĐTĐL/2011-T/29/HĐ).

References

- Bonafonte MT, Smith LM, Mead JR. 2000. A 23-kDa recombinant antigen of *Cryptosporidium parvum* induces a cellular immune response on in vitro stimulated spleen and mesenteric lymph node cells from infected mice. *Experimental Parasitology*, 96(1): 32-41.
- Borad AJ, Allison GM, Wang D, Ahmed S, Karim MM, Kane AV, Moy J, Hibberd PL, Rao Ajjampur SS, Kang G, Calderwood SB, Ryan ET, Naumova E, Khan WA, Ward HD. 2012. Systemic antibody responses to the immunodominant p23 antigen and p23 polymorphisms in children with cryptosporidiosis in Bangladesh. *The American Journal of Tropical Medicine and Hygiene*, 86(2): 214-222.
- Inpankaew T, Jittapalapong S, Phasuk J, Pinyopanuwut N, Chimnoi W, Kengradomkit C, Sunanta C, Zhang G, Aboje GO, Nishikawa Y, Igarashi I, Xuan X. 2009. Seroprevalence of *Cryptosporidium parvum* infection of dairy cows in three northern provinces of Thailand determined by enzyme-linked immunosorbent assay using recombinant antigen CpP23. *Onderstepoort J. Vet. Res.*, 76(2): 161-165.
- Jenkins MC, Fayer R. 1995. Cloning and expression of cDNA encoding an antigenic *Cryptosporidium parvum* protein. *Molecular and Biochemical Parasitology*, 71(1): 149-152.
- Klein D, Kern A, Lapan G, Benetka V, Möstl K, Hassl A, Baumgartner W. 2009. Evaluation of rapid assays for the detection of bovine coronavirus, rotavirus A and *Cryptosporidium parvum* in faecal samples of calves. *The Veterinary Journal*, 182(3): 484-486.
- Laxer MA, Timblin BK, Patel RJ. 1991. DNA sequences for the specific detection of *Cryptosporidium parvum* by the Polymerase Chain Reaction. *The American Journal of Tropical Medicine and Hygiene*, 45(6): 688-694.
- Liu K, Zai D, Zhang D, Wei Q, Han G, Gao H, Huang B. 2010. Divalent Cp15-23 vaccine enhances immune responses and protection against *Cryptosporidium parvum* infection. *Parasite Immunology*, 32(5): 335-344.
- Luginbühl A, Reitt K, Metzler A, Kollbrunner M, Corboz L, Deplazes P. 2005. Feldstudie zu Prävalenz und Diagnostik von Durchfallerregern beim neonaten Kalb im Einzugsgebiet einer schweizerischen Nutztierpraxis. *Schweizer Archiv für Tierheilkunde*, 147(6): 245-252.
- Makrides SC. 1996. Strategies for achieving high-level expression of genes in *Escherichia coli*. *Microbiol. Rev.*, 60: 512-538.
- McNabb SJ, Hensel DM, Welch DF, Heijbel H, McKee GL, Istre GR. 1985. Comparison of sedimentation and flotation techniques for identification of *Cryptosporidium* sp. oocysts in a large outbreak of human diarrhea. *J. Clin. Microbiol.*, 22(4): 587-589.
- O'Donoghue PJ. 1995. *Cryptosporidium* and cryptosporidiosis in man and animals. *International Journal for Parasitology*, 25(2): 139-195.
- Perryman LE, Jasmer DP, Riggs MW, Bohnet SG, McGuire TC, Arrowood MJ. 1996. A cloned gene of *Cryptosporidium parvum* encodes neutralization-sensitive epitopes. *Molecular and Biochemical Parasitology*, 80(2): 137-147.
- Perryman LE, Kapil SJ, Jones ML, Hunt EL. 1999. Protection of calves against cryptosporidiosis with immune bovine colostrum induced by a *Cryptosporidium parvum* recombinant protein. *Vaccine*, 17: 2142-2149.
- Priest JW, Kwon JP, Moss DM, Roberts JM, Arrowood MJ, Dworkin MS, Juranek DD, Lammie PJ. 1999. Detection by enzyme immunoassay of serum immunoglobulin G antibodies that recognize specific *Cryptosporidium parvum* antigens. *Journal of Clinical Microbiology*, 37(5): 1385-1392.
- Shahbazi P, Shayan P, Ebrahimzadeh E, Rahbari S. 2009. Specific egg yolk antibody against recombinant *Cryptosporidium parvum* P23 protein. *Iranian Journal of Parasitology*, 4(3): 15-24.
- Shayan P, Ebrahimzadeh E, Mokhber-Dezfouli M-R, Rahbari S. 2008. Recombinant *Cryptosporidium parvum* p23 as a target for the detection of *Cryptosporidium* specific antibody in calf sera. *Parasitology Research*, 103: 1207-1211.
- Shirafuji H, Xuan X, Kimata I, Takashima Y, Fukumoto S, Otsuka H, Nagasawa H, Suzuki H. 2005. Expression of P23 of *Cryptosporidium parvum* in *Toxoplasma gondii* and evaluation of its protective effects. *J. Parasitol.*, 91(2): 476-479.
- Wang C, He H, Duan M. 2009. Development and evaluation of a recombinant CP23 antigen-based ELISA for serodiagnosis of *Cryptosporidium parvum*. *Experimental Parasitology*, 121(2): 157-162.
- World Organisation for Animal Health. 2008. *Cryptosporidiosis. Manual of Diagnostic Tests and Vaccines for Terrestrial Animals*, vol. 6. Office international des epizooties, Paris, France.